Neuroimmunological and Neuroenergetic Aspects in Exercise-Induced Fatigue

Sebastian Proschinger¹ and Jens Freese²

- ¹ German Sport University Cologne, Department for Molecular and Cellular Sports Medicine, Am Sportpark Müngersdorf 6, 50933 Cologne, Germany
- ² Dr. Freese Institute for Exercise & Nutritional Immunology, Josef-Lammerting-Allee 7-13, 50933 Cologne, Germany

Abbreviations

ADO	—	adenosine
ATP	_	adenosine triphosphate
BBB	_	blood-brain barrier
CNS	_	central nervous system
CVO	_	circumventricular organs
DAMP	—	danger-associated molecular pattern
GABA	_	γ-aminobutyric acid
IL-1	_	Interleukin 1
IL-6	_	Interleukin 6
LPS	_	lipopolysaccharide
PAMP	_	pathogen-associated molecular pattern
RNS	_	reactive nitrogen species
ROS	_	reactive oxygen species
S100	_	S100 calcium-binding protein
TNF	_	tumor necrosis factor-alpha
TLR	_	Toll-like receptor
VO_{2max}	_	maximal oxygen consumption
5-HT	_	5-hydroxytryptamine

Index

- 1. Introduction
- 2. Peripheral and central fatigue
 - 2.1 Lactate accumulation
 - 2.2 Neurotransmission
 - 2.3 Cytokines
- 3. Systemic inflammatory response during exercise muscle damage, leukocytosis and endotoxemia
- 4. Communication interfaces between periphery and central nervous system
- 5. Neuroinflammation and fatigue
- 6. Energetic Regulation is there a selfish immune system in the brain?
- 7. Purinergic regulation of neuroinflammation and neurotransmission in the basal ganglia
- 8. Neuroinflammation-induced energy reallocation during exercise a new paradigm?

Sebastian Proschinger¹, s.proschinger@gmx.net Jens Freese², info@dr-freese.com

ABSTRACT

Feelings of fatigue not only occur in chronic and acute disease states, but also during prolonged strenuous exercise as a symptom of exhaustion. The underlying mechanisms of fatigue in diseases seem to rely on neuroinflammatory pathways. These pathways are interesting to understand exerciseinduced fatigue regarding immune system to brain signaling and effects of cerebral cytokines. Activation of the immune system incurs a high-energy cost, also in the brain. In consequence immune cells have high energetic priority over other tissues, such as neurons. A neuronal inactivation and corresponding changes in neurotransmission can also be induced by end products of ATP metabolism and elicit feelings of fatigue in diseases and after intensive and prolonged exercise bouts. Since there are no existing models of exercise-induced fatigue that specifically address interactions between neuroimmunologic mechanisms and neuroenergetics, this article is combining scientific evidence across a broad range of disciplines in order to propose an inflammation- and energy-based model for exercise-induced fatigue.

Keywords: exercise-induced fatigue, neuroinflammation, neuroenergetics, adenosine, cytokines.

1. Introduction

To study exercise-induced fatigue for many years, priority was given to muscles over the brain as a regulatory factor. Already in 1915, Alessandro Mosso postulated that both, the will (central component) and the muscular work (peripheral component), have to be taken into account when considering the resulting impairment of exercise performance. Mosso distinguished the diminution of the muscular force and the sensation of fatigue (1). As a result of the upcoming knowledge of the bi- and multidirectionality of biological systems, the paradigm shifted to the inclusion of cerebral processes in order to guarantee homeostasis in all systems during exercise by modulating athlete's behavior (2,3).

Since proinflammatory cytokines induce changes in behavior during acute infection by provoking feelings of fatigue (4,5), it is reasonable that the remarkable rise in circulating proinflammatory signals during prolonged strenuous exercise (6) may also contribute to exercise-induced fatigue. In this regard, the neuromodulatory properties of myogenic/neuronal Interleukin 6 (IL-6) and cerebral immune cell-derived Interleukin 1 (IL-1) have recently been discussed as major factors in exercise-induced fatigue (7,8).

^{*} Corresponding author:

According to the selfish immune system theory (9), high synthesis rates of cytokines indicate high energy turnover of immune cells and with that, higher energetic needs of those. In the case of an increasing brain macrophage activity, energy substrates may be shifted away from neurons to these immune cells to maintain their activity (8,10). Because decreasing neuronal activity seems to induce feelings of fatigue also during exercise, a compromised energy provision to neurons due to increasing brain immune cell activity could account for the decline in exercise performance (11,12).

The initially increasing neuronal and glial energy turnover during prolonged strenuous exercise (11,12) may favor the generation of the nucleoside adenosine (ADO) (13), which negatively mediates exercise performance in a concentrationdependent manner by modulating dopamine neurotransmission in the basal ganglia (14,15).

Here, we propose that (neuro-)immunological mechanisms influence neuroenergetics, with both proinflammatory signals and end products of energy turnover inducing feelings of fatigue during prolonged strenuous exercise and ultimately provoking exercise termination.

2. Peripheral and central fatigue

Already in the late nineteenth century, the physiologist Angelo Mosso postulated that "muscular fatigue also is at bottom an exhaustion of the nervous system" (2). In the context of exercise-induced fatigue, central or supraspinal fatigue appears to originate in regions of the brain and is defined as the inability of the CNS to drive motor neurons efficiently during the performance of intermittent or prolonged aerobic exercise (16), whereas peripheral or muscle fatigue is the result of biochemical changes in the exercising limb muscles (17).

2.1 Lactate accumulation

According to the lactate theory of exercise fatigue, the exercising muscles stop working due to a massive intracellular lactate accumulation as a consequence of an insufficient supply of oxygen and the upregulation of the muscle cell's anaerobic metabolism (17).

However recent findings challenge the correctness of the lactate theory (Robergs, 2004) and emphasize the significance of lactate as energy substrate in other metabolic processes (18,19). Via intracellular monocarboxylate transport proteins, lactate is used as an additional energy substrate both by contracting and adjacent inactive muscle fibers. During strenuous exercise, a reciprocal brain-muscle energy exchange occurs in which the brain favors muscle-derived lactate in order to provide enough circulating glucose to type-2 muscle fibers as its primary energy substrate (20-22).

The energetic capacity of exercising muscles does not decrease significantly to promote peripheral fatigue, since muscles are still capable to generate power at exhaustion (23). Because neither lactate accumulation in exercising muscles nor associated muscle acidification cause peripheral fatigue (23), these findings underline the assumption of exercise termination forced by central mechanisms.

2.2 Neurotransmission

Neurotransmission of monoamines plays a crucial role in exercise-induced fatigue. The central fatigue hypothesis postulated by Newsholme et al. (24) states that exercise-induced synthesis of cerebral serotonin (5-HT) provokes the onset of fatigue symptoms. Since 5-HT can not cross the blood-brain barrier (BBB), brain cells rely on the uptake of tryptophan as its precursor. Animal studies (25) have shown that tryptophan injections in the cerebral ventricle of rats were associated with the onset of exercise-induced fatigue, while inhibition of the conversion of tryptophan to 5-HT could improve running time to fatigue. However, others have proven a reduction in plasma tryptophan in humans after exhaustive aerobic exercise (26), which seems contradictory to the aforementioned findings. Strasser et al. conclude that there is limited availability of tryptophan for 5-HT biosynthesis in the brain after the enzymatic conversion to kynurenine in the periphery.

Recent findings provide evidence that dopaminergic neurotransmission in striatopallidal neurons increases exercise performance by maintaining motivation and motor regulation (27,28). A blockage of central dopaminergic D1/D2 receptors results in a significant decrease in endurance performance and maximal oxygen uptake (29).

2.3 Cytokines

Many systemic inflammatory and neuroinflammatory disorders, i.e. chronic fatigue syndrome (CFS), depression or multiple sclerosis, are frequently accompanied by high amounts of circulating cytokines and a persistent state of mental and physical fatigue (30). Neuroimaging studies have suggested the presence of neuroinflammation in the midbrain of CFS patients (31). Furthermore, CFS patients achieve volitional exhaustion significantly faster and consistently report a higher rate of perceived exertion during an exercise task, assuming that CFS, in part, is mediated centrally (32). Chronic fatigue in athletes suffering from overtraining/athlete burnout may also result from circulating proinflammatory cytokines and a neuroinflammatory state (33,34).

Vargas & Marino (35) proposed a neuroinflammatory model for acute fatigue during exercise. The authors suppose a potential interaction between cytokine release during prolonged strenuous exercise and their effects on afferent feedback signalling to the brain that might lead to feelings of fatigue. In particular, the extraordinary increase in plasma IL-6 concentration is proposed to be a major fatigue-inducing factor due to its receptor-mediated signal transduction in neuronal afferents and circumventricular organs (CVO).

Already in 2000, the influence of muscle-derived IL-6 was considered to play an important role in the development of central fatigue (36). Subcutaneous administration of a low dose of recombinant IL-6 to athletes increase their sensation of fatigue at rest and significantly impairs athletic performance during a 10-km running time trial (37). Because of its autocrine, paracrine or endocrine effects, muscle-derived IL-6 may also function as an energy sensor and a hormone-like molecule that increases energy substrate mobilization (38-40), possibly by an intensity-dependent upregulation of cortisol (41,42). Therefore, high IL-6 levels could represent the need for energy substrates.

After an eccentric exercise bout, the concentration of IL-1 increases significantly in rat brain regions responsible for movement, coordination, motivation, perception of effort, and pain. Its levels correlate significantly with both post-exercise delayed recovery and decreased performance in a subsequent task (43). Further, intracerebroventricular injection of IL-1 significantly decreased wheel running activity in uphill running mice, whereas IL-1ra improved wheel running in downhill running mice (44). Another study identified perivascular and meningeal macrophages as the major producer of brain IL-1 during exercise (8).

There is vast evidence that microglia, another mononuclear phagocytic cell type in the CNS and the main actor in neuroin-flammation, synthesize both IL-1 and TNF in high amounts after activation. Furthermore, the decrease in symptoms of depression and fatigue is accompanied by a reduced TNF secretion in the CNS through modulation of neuroinflammation (31,45,46).

3. Systemic inflammatory response during exercise – muscle damage, leukocytosis and endotoxemia

Via the production of IL-6 and reactive oxygen species (ROS), both exercise-induced muscle damage (47,48) and the intensity-dependent rise in circulating T-lymphocytes and neutrophils (49,50) significantly contribute to the exercise-induced systemic inflammation (51,52). The rise in serum neopterin during exhaustive aerobic exercise suggests an increased activation of peripheral macrophages (26). However, results from Ostrowski et al. (53) reveal an increase of the anti-inflammatory cytokines IL-10, IL-1 receptor antagonist and soluble TNF receptors during and after strenuous exercise, possibly due to the massive increase in IL-6 (41,54).

Lymphocyte-derived extracellular heat shock proteins are known to increase during high-load exercise and are further proposed to promote fatigue sensation via marked influence on motor neurons and deeper structures of the CNS (55). These molecules also promote inflammation by acting as a danger signal from the immune system. Bårdsen et al. (56) suggest that the significant increase in extracellular heat shock proteins in CFS patients might signal to the brain and contribute to the state of fatigue.

The observation that prolonged strenuous exercise favors a systemic inflammatory state was discussed by John Marshall, assuming that the exercise-induced increase in intestinal permeability and lipopolysaccharide (LPS)-induced endotoxemia may be the underlying cause (57). LPS is a gut-derived proinflammatory fragment of the outer membrane of gram-negative bacteria and a pathogen-associated molecular pattern (PAMP). Pals et al. (58) showed that the degree of the intestinal permeability depends mainly on exercise intensity and correlates with body core temperature. In fact, human studies show that the severity of endotoxemia seems highly dependent on the environmental temperature (59,60), but also on the composition of the gut microbiota (61). In this regard, the supplementation of probiotics over a period of 4 weeks displays a tendency to decreasing intestinal permeability and reducing LPS in the bloodstream (62). After an ultramarathon, 81% of the participants showed plasma LPS levels > 0.1 ng/ml (endotoxic), while 2% even had a plasma concentration of 1 ng/ml (potentially lethal) despite moderate environmental temperatures ($20,3^{\circ}C-22,3^{\circ}C$) (63). Both exercise-induced functional splanchnic hypoperfusion and translocation of LPS are damaging the protein-barrier complex between enterocyte membranes via temperaturedependent and immune-mediated mechanisms (64-66). This contributes to an endotoxic state.

4. Communication interfaces between periphery and central nervous system

A systemic inflammatory response has been shown to affect the activity of immune cells in the brain. The growing importance of the bidirectionality between the periphery and the central nervous system (CNS) and the impact of neuroimmunomodulatory mechanisms (67) puts the interplay of endocrine, neuronal and immunological mechanisms in the forefront of exercise regulation (3). Due to acute or chronic immune stressors, dysregulation at periphery-CNS interfaces, i.e. the BBB, CVO, and afferent nerve fibres (68), is associated with pathological conditions in which fatigue is a common feature (69). As prolonged strenuous exercise represents a huge physiological stressor accompanied by immune activation, interface-specific cells could get regulated in order to induce systemic adaption and maintain homeostasis in all systems during exercise (2,70).

Some cytokines use specific mechanisms to access the brain parenchyma by bypassing its saturable transport mechanisms (71). The serum level of the S100 calcium-binding protein (S100) which provides information about the severity of the BBB's permeability, increases during strenuous exercise (72). Both duration and intensity of an exercise bout (73) and game-related activities or events (74) seem to determine the rise in S100 plasma concentration. Furthermore, S100 is the most frequently assessed biomarker in studies investigating sport-related concussion which is known to induce BBB disruption (75). According to the severity of concussion, the post-injury neuroinflammatory state promotes metabolic dysfunction and neuronal impairment (76), often followed by persistent feelings of fatigue, without regard of traumatic brain injury severity (77,78). A correlation between the onset of exercise-induced fatigue and the number or magnitude of impacts to the head is possible, but experimental data are lacking.

Although LPS is able to alter transport rates for many peptides across the BBB (79), LPS acts on receptors outside the BBB rather than directly on BBB's structures to modulate its integrity (80,81). Peripheral administration of subseptic doses of LPS initiates the synthesis of IL-1 and tumor necrosis factor-alpha (TNF) messenger RNA at the CVO, but not at the BBB (82). Since plasma LPS concentration can rise significantly during prolonged strenuous exercise (63), CVO could play a decisive role in neuroimmunological modulation. Recent studies show that communication between peripheral immune cells and brain structures predominantly occurs at the sensory CVO (83). Their unique structure enables them to monitor and transmit blood- and cerebrospinal fluid-derived information from circulating substances that do not readily cross the BBB.

During a systemic inflammatory response, concentrations of the IL-6 receptor and the IL-6 signal transducer glycoprotein 130 are highest in the sensory CVO. The synthesis rate of both increase significantly in accordance with circulating IL-6 (84), thereby enforcing its neuroimmunomodulatory properties. The huge rise in serum IL-6 during prolonged strenuous exercise may increase levels of soluble IL-6 receptor and glycoprotein 130 in the sensory CVO. A systemic inflammatory response upregulates IL-1 receptor and TLR (Toll-like receptor) 4 in the sensory CVO as well, both changing the activity of neurons and inducing gene expression of proinflammatory cytokines (85-87). The IL-1 receptor and TLR4 is expressed by microglia and by brain macrophages. After a single systemic administration of LPS, microglia show increased proliferation in the sensory CVO compared with other regions of the brain (88), presumably compensating for the lack of a protecting BBB.

Receptors for cytokines and LPS are also expressed at the terminal nerve endings of the vagus nerve, suggesting a crucial role in immunomodulation and sickness behavior via signalling from nucleus tractus solitarius to brainstem, hypothalamus and higher brain centers (89-91). Once these receptors become activated, the vagus nerve is stimulated in a dosedependent relationship (92,93).

Since the afferent activity of the hepatic vagus nerve seems to contribute to the orchestration of the metabolic and hormonal responses to exercise, cytokine-induced stimulation of the vagus nerve could influence exercise performance in a dose-dependent manner (94). Similarly, activation of glial cells in the spinal cords of mice during eccentric exercise alters their gene expression due to the emerging skeletal muscle inflammation (95), provoking exercise-induced muscle hyperalgesia by IL-6 signalling on primary afferent nociceptors (96). Enhanced glial cytokine synthesis in the spinal cord is also shown during acute and chronic pain states and in inflammatory muscle disease (97,98) with fatigue and pain pathways being quite similar regarding cytokine signalling (99).

5. Neuroinflammation and fatigue

Since the perception of fatigue as a hallmark of sickness behavior seems to be cytokine-driven (4,5), fatigue is widespread in people suffering from neurodegenerative and chronic inflammatory diseases (30). Both direct and indirect measurement methods revealed an increased intestinal permeability, higher circulating LPS levels and a region-specific rise in neuroinflammation (100-104). Therefore, a causal relationship between intestinal permeability, neuroinflammation and the perception of fatigue is reasonable.

Rats exposed to either an immunological or a physical stressor show symptoms of sickness behavior in a time-dependent manner. However, when IL-1 receptor antagonist is injected intracerebroventricularly prior to the physical stress exposure, symptoms do not appear (105). Indeed, IL-1 and IL-6 may function as immunological correlates of human sickness behavior. During infection, levels of IL-1 and IL-6 spontaneously released from peripheral blood mononuclear cell cultures were consistently correlated with reported manifestations of acute sickness behavior including fever, malaise, pain, fatigue, mood and poor concentration (106).

An animal study showed that the administration of antiinflammatory omega-3 fatty acids significantly inhibit LPSinduced neuroinflammation in the prefrontal cortex, hippocampus and hypothalamus and reverses depression-like behavior (46). Moreover, supplementation of the omega-3 fatty acid eicosapentaenoic acid in the course of 16 weeks promotes symptom remission and structural brain changes in patients with CFS (107).

6. Energetic regulation – is there a selfish immune system in the brain?

From an ecoimmunological point of view, an acute inflammatory response is metabolically extremely costly according to its allostatic load (108). As allostasis is an evolutionarily conserved and energy-intensive response to resume local homeostasis, the allostatic load indicates the severity of the homeostatic disruption (109). Based on in vitro O2-consumption rates (24), activated macrophages turn over ATP ten times faster per minute compared to the inactivated state. The favored aerobic glycolysis of activated immune cells makes glucose their primary energy substrate (110), using strategies to redistribute energy to themselves to keep their metabolism running (9). New insights indicate that these characteristics can also be observed in microglia depending on their polarization state (10,88,111).

Assuming that brain macrophages become overactive during prolonged strenuous exercise (8), their energy needs could reduce energy provision to neurons, thereby promoting the occurrence of fatigue symptoms. In patients with tuberculous meningitis, the infection with Mycobacterium tuberculosis represents a huge allostatic load indicated (112). The infection is accompanied by microglial activation and the allocation of astrocytic lactate to microglia via astrocyte-microglia lactate shuttles, thereby providing an adequate energy supply activated microglia. As a result, the allocation of lactate to neurons decreases significantly, which leads to neuron inactivation (10). Similarly, when lactate shuttling from astrocytes to neurons decreases during strenuous exercise, neurons are not able to maintain their metabolism (11,113). In consequence, exercise performance declines.

Acute bouts of strenuous exercise mobilize highly differentiated T-cells from peripheral tissues into the blood stream (49,114) referring to exercise-induced leukocyte demargination (115). Since a high differentiation level is associated with decreased mitochondrial content and function, these immune cells mainly rely on the glucose-consuming anaerobic metabolism (116). The trafficking rate depends on the aerobic fitness level with untrained people showing higher redistribution of these energy consuming immune cells into the blood stream (117,118), potentially contributing to the earlier onset of exercise-induced fatigue in this population.

A high energy turnover induces ATP breakdown to ADO. ADO is secreted by ATP-depleted tissues or is extracellularly generated from ATP, which is released from metabolically stressed cells (119). In a Drosophila infection model, ADO induces energy reallocation by enhancing uptake of glucose in immune cells at the expense of other glucose-dependent tissues, including the brain (120). Consequently, ADO is considered being a signalling molecule whose effects could increase fatigue in relation to the energetic demand of activated immune cells (121). It is important to note that ADO regulatory and signaling network in Drosophila is similar to mammalian systems (121). Since high levels of ADO accumulate in the brain after prolonged strenuous exercise (13), it is reasonable that there could be similar mechanisms of action.

7. Purinergic regulation of neuroinflammation and neurotransmission in the basal ganglia

New insights into mechanisms of action of purines in the CNS with respect to neuroinflammatory processes and behavioral regulation emphasize their neuromodulatory effects, although most results are from animal studies (122). A rise in extracellular ADO favors neuroinflammatory signalling through upregulation of the high-affinity A_{2A} adenosine receptor (123). As high amounts of extracellular ATP are considered to be evolutionarily conserved danger-associated molecular pattern (DAMP) (124), it initiates inflammation via stimulation of the TLR4-dependant cytosolic inflammasome in microglia (125). While both ADO and ATP are able to enhance the production of IL-1 (126), IL-1 in turn promotes ATP and ADO release from neurons (127). Experimental data in mice suggest a potentiation of nitric oxide release by activated microglia after interacting with the A2A adenosine receptor, thereby increasing ROS and reactive nitrogen species (RNS) production (128-130). In addition, the stimulation of the ATP-purinoceptors P2X7R and P2X4R favors synthesis of IL-6 and TNF, what further promotes neuroinflammation (131).

ADO directly influences behavior by decreasing dopaminergic neurotransmission through conformational changes of D2R binding sites at a shared A_{2A}/D_2 - and $A_{2A}/D_2/mGlu_5$ receptor complex on rat striatopallidal GABA neurons (15,132,133). As dopamine is an important neurotransmitter in exercise regulation, ADO may negatively influence exercise performance in rats (134). In contrast, the ADO antagonist caffeine delays run time to fatigue in rats by 52%, presumably by increasing dopamine release through an antagonism at the A_1 and A_{2A} adenosine receptors in the striatum, the nucleus accumbens and the nucleus caudatus (135) or the preoptic area and the anterior hypothalamus (136). However, no effect of caffeine on exercise performance was seen in humans exercising in high ambient temperature (137).

8. Neuroinflammation-induced energy reallocation during exercise – a new paradigm?

Not only exercise-induced muscle damage, endotoxemia and leukocytosis contribute to the systemic inflammatory response in exhausted athletes, but also the release of ROS/RNS and, to a lower extent, cytokine-dependent apoptosis of leukocytes and neutrophils immediately after prolonged strenuous exercise (138,139). Although circulating lymphocytic subpopulations contain a high antioxidant capacity (140), it is conceivable that leukocytes whose capacity has already been exhausted during prolonged strenuous exercise could undergo apoptosis even before exercise termination. Cells that are not immediately phagocytosed after apoptosis become "leaky" (secondary necrosis). They release DAMPs and stimulate a host response by secreting more proinflammatory signals (141).

Exercise-induced rise in serum LPS concentration may induce changes at the BBB and favors microglia proliferation at the CVO, thereby inducing neuroinflammation (80,88). If gutderived LPS accumulates in the liver by overwhelming the capability of the liver's reticulo-endothelial system (63), the resulting stimulation of Kupffer cells may force the secretion of cytokines. Binding of LPS and IL-1 to receptors on terminal nerve endings of the hepatic vagus nerve may activate microglia (69,142).

There is some evidence that IL-6 acts as a major factor and is contributing to exercise-induced fatigue (7,36,37). Results from prolonged (marathon) and highly prolonged (spartathlon) endurance exercise show a 128-fold and respectively 8000-fold increase in IL-6 plasma levels, peaking at exercise termination and rapidly normalizing afterwards (53,143). This outcome may support the fatigue-inducing character of IL-6 instead of being a proinflammatory cytokine in the context of exercise. However, as energy availability declines drastically due to the physical strain in such events, muscle-derived IL-6 may also work in its hormone-like fashion by increasing energy substrate mobilization (38-40).

Since increased neuronal metabolism alters microglia functioning, neurons can be regarded as key immune modulators in the brain (144). As neuronal metabolism and extracellular levels of 'neuron-microglia signalling factors' rise, they function as "On" signals (Fig. 1: right box, dark blue arrows) by recruiting microglia which then support the neuron's metabolism (Fig. 1: microglia-astrocyte-neuron lactate shuttles = right box, purple arrow). Already before, astrocytes begin to serve the energy needs of the neurons through cellular lactate transfer (Fig. 1: astrocyte-neuron lactate shuttles = light blue arrow).

The rise in extracellular ADO due to the high glial and neuronal ATP turnover may increase astrocyte proliferation and activation (145). The significant increase in brain ADO during strenuous exercise (13), could, therefore, aim to enhance astrocytic lactate production to supply the cells in need (Fig. 1). Furthermore, cerebral ADO modulates BBB permeability through stimulation of endothelial $A_{2A}R$ and $A_{1}R$ (146). An enhanced uptake of blood lactate may be the consequence, as

a moderately increased permeability of the BBB is regarded as a functional mechanism during exercise by serving neuronal metabolism (27). Marked changes, however, could may limit the individual's capacity to perform optimally by allowing the accumulation of unwanted substances in the CNS (27).

Because almost all metabolic processes show a dose-response relationship during stress exposure with both beneficial and detrimental outcomes (147), exercise above a certain threshold can cause mal-adaptations as well (148). Regarding prolonged strenuous exercise, the exercise-related dose response induces an inflammatory state (Fig. 1) and may also provoke an acute neuroinflammatory response (8) due to the high allostatic load on brain cell metabolism. However, experimental data are lacking to make clear conclusions about brain metabolism during exercise and its relation to neuroinflammation. But to integrate the existing knowledge about exercise-related dose response into the concept of neuroinflammation, we propose a model of continuum in which the astrocyte-neuron lactate shuttle expands to the microglia-astrocyte-neuron lactate shuttle (149) when energy demand of neurons increase during exercise (Fig. 1, right box). Both, the intensity-dependent systemic inflammatory response and brain cell-derived purines may switch the microglial phenotype from the M2/antiinflammatory form to the M1/proinflammatory form, thereby making them more "energy-craving". That is followed by a step-by-step inactivation of neurons through astrocytemicroglia lactate shuttles (10) (Fig. 1: right box, red arrow).



Figure 1: Hypothetical integrative model showing how neuroimmunological and neuroenergetic mechanisms induce feelings of fatigue during prolonged strenuous exercise, ultimately provoking exercise termination.

Strenuous exercise favors exercise-induced muscle damage, gut-derived LPS translocation and immune cell expansion (leukocytosis) [1]. At the same time, the exercise-induced and intensity-dependent increase in neuronal metabolism favors the release of neuronal 'On'-signals, which induce lactate transfer from glial cells to neurons, beginning with the astrocyte-neuron lactate shuttle [2] and extending to the microglia-astrocyte-neuron lactate shuttle in order to serve the increasing energy needs of the neurons [3]. Microglial autoactivation through microglia-derived IL-1 β and extracellular ATP may promote a switch to the M1/proinflammatory form. As strenuous exercise continues, that leads to a proinflammatory state characterized by high circulating amounts of LPS, DAMPs (e.g. HSPs), IL-6 and ROS-damaged immune cells. These proinflammatory signals act either on the BBB, CVOs and VN which then signal to the CNS or they act directly on the CNS by passing the BBB or CVOs [4]. In doing so, these signals may stimulate microglia/brain macrophages through TLR-4 and IL-1 β receptors [5] which then continuously shut down the lactate transfer from astrocytes to neurons through endo- and ectonucleases favor accumulation of extracellular adenosine that impairs dopaminergic neurotransmission by acting on the A2A/D2/mGLU5 receptor complex on striatal neurons [7]. A possible contribution of IL-1 β to a denosine signalling may further enhance the down regulation of dopaminergic neurotransmission. The resulting neuronal inactivation [8] leads to a decline in cognition, vigilance and neuromuscular activation, ultimately inducing exercise-induced fatigue [9].

 A_{2A} : adenosine A_{2A} receptor; D_2 : dopamine D_2 receptor; mGLU₅: metabotropic glutamate receptor 5; IL-1 β : interleukin 1 β LPS: lipopolysaccharide; IL-6: interleukin 6; DAMP: danger-associated molecular patterns; BBB: blood-brain barrier; CVO: circumventricular organs; VN: vagus nerve; AMP: adenosine monophosphate; ATP: adenosine triphosphate; TLR-4: toll-like receptor 4; HSP: heat shock protein; CNS: central nervous system.

This microglial polarization is often accompanied by a shift from oxidative phosphorylation to aerobic glycolysis for energy production due to increasing concentrations of nitric oxide by inducible nitric oxide synthetase which reversibly inhibits mitochondrial respiration (111). With that, ROS and RNS production is increased which, in turn, activates downstream signaling pathways resulting in the up-regulation of a variety of proinflammatory proteins and more ROS/RNS.

Whether there is a similar mechanism of energy reallocation from neurons to activated microglia/brain macrophages during non-infectious stress is unknown. However, haemodynamically stressed microglia express monocarboxylate transporter-1 and -2 (150), which may enable them to utilize astrocytic glycogen-derived lactate. Since there is remarkable cerebral haemodynamic stress during prolonged strenuous exercise (151), expression of monocarboxylate transporters may promote the uptake of astrocytic lactate in microglia or brain macrophages.

Although the amount of LPS crossing the BBB is low (80,81), some athletes show plasma concentration of 1 ng/ml after an ultramarathon (63). If LPS crosses the BBB at that concentration is unknown, but conceivable since the BBB becomes leakier during strenuous exercise. As LPS-TLR4 interactions resemble proinflammatory pathways induced by Lipoarabinomannan, the major cell wall component of mycobacterium tuberculosis (152,153), high amounts of LPS in the brain may be able to induce the expression of astrocyte-microglia lactate shuttles. Further, cerebral DAMPs may promote astrocytemicroglia lactate shuttles in a similar fashion by triggering the microglial TLR4 (Fig. 1).

Heck et al. (55) propose that the exercise-induced increase in circulating levels of extracellular 70-kDa heat shock proteins from lymphocytes promote fatigue via marked influence on motor neurons and deeper structures of the CNS. Although specific receptors for heat shock proteins in brain tissue have not been identified yet, their ability to induce proinflammatory signalling in TLR4/2-expressing cells is well established (154,155).

Because lactate does not accumulate in cerebrospinal fluid after an exhaustive exercise task (156), unlike during tuberculous meningitis (157), we do not know whether it is appropriate to think of the astrocyte-microglia lactate shuttles as a relevant mechanism in exercise-induced fatigue. Further it is unknown whether extracellular ADO reallocates energy substrates to demanding cerebral immune cells and thereby shutting down the less relevant neuronal metabolism as shown in a Drosophila infection model on the peripheral level. Extracellular ADO definitively compromises exercise performance in animals due to its inhibitory effect on dopaminergic neurotransmission (134,135). To connect the potential fatigueinducing property of ADO, Hanff et al. (158) assume that it plays an important role in the induction of sickness behavior via the A2A/D2/mGLU5-receptor-complex (Fig. 1). In fact, LPS-induced swim deficits is reversed by systemic administration of an A2A receptor antagonist (159). A similar receptor-ligand interaction appears to be relevant in the induction of sleep (160). The stimulation of A2AR and mGLU5R inhibits the activity in vigilance-regulating brain areas by presynaptic inhibition, postsynaptic hyperpolarization and amplifying GABAergic projections (161,162). Increased dopamine release in the ventral tegmental area reduces the inhibitory activity in the nucleus accumbens and is promoting vigilance.

Dopaminergic neurotransmission in the substantia nigra pars compacta inhibits neuroinflammation by activating astrocytic D₂-receptors (163). Based on the assumption that IL-1 may contribute to motivational and vigilance regulation via an important interaction with ADO signalling in the CNS, i.e. activation of A2A receptors in the striatum (158) (Fig. 1), the attenuated dopamin-induced anti-inflammatory effect could promote synthesis of IL-1. Both, inflammation- and exerciseinduced peripheral hyperammonaemia promote cerebral synthesis of ADO (164,165), which may force exercise-induced fatigue by altering cognition (165). The increasing impairment of the fronto-striatal network down-regulates cognition and motivation, which makes exercise termination rather a relative than an absolute event due to the athlete's volitional and forced conscious decision (see Fig. 1) (3,166,167). The impact of peripheral cytokine signalling and central microglia/brain macrophage activation on this fronto-striatal network should be taken into account (69).

CONCLUSION

Exercise-induced fatigue does not emerge from a single peripheral or central mechanism, but rather result of a synergistic effect of various mechanisms involving both peripheral and central aspects. As an evolutionary conserved protective mechanism, neuron inactivation and the concomitant increase in feelings of fatigue are extremely useful to maintain systemic homeostasis at all bodily levels, also during exercise. If the immune system is even selfish in the brain, microglia/brain macrophage-derived extracellular ADO could mediate the metabolic switch and energy reallocation, thereby inducing neuron inactivation, feelings of fatigue and ultimately exercise termination. Due to the impact of IL-1 on feelings of fatigue and behavior modulation, the synthesis of IL-1 from perivascular and meningeal macrophages during strenuous exercise has to be considered when approaching the complexity of exercise-induced fatigue. Changes in cerebral haemodynamics are not investigated in this article but should be subject of further studies about the regulation of exercise performance. In order to get deeper insights into the brain metabolism during prolonged strenuous exercise and its relation to neuroinflammation, the hormesis-like dose response of brain macrophage activation during exercise should be investigated in future studies.

As presented here, the majority of aspects concerning neuroimmune-neuroenergetic interactions in sports performance are not very well established and need to be evaluated in the future. Therefore, it is inevitable to improve interdisciplinary research in this field.

REFERENCES

- Matthews, G., Hancock, P. A., & Desmond, P. A. The handbook of operator fatigue. CRC Press, 2018.
- 2. Noakes TD. Fatigue is a Brain-Derived Emotion that Regulates the Exercise Behavior to Ensure the Protection of Whole Body Homeostasis. Front Physiol 3: 82, 2012.
- Kayser B. Exercise starts and ends in the brain. Eur J Appl Physiol 90: 411-419, 2003.
- Dantzer R, O'Connor JC, Freund GG, Johnson RW, Kelley KW. From inflammation to sickness and depression: when the immune system subjugates the brain. Nat Rev Neurosci 9: 46-56, 2008.
- 5. Hart BL. Biological basis of the behavior of sick animals. Neurosci Biobehav Rev 12: 123-137, 1988.
- Fehrenbach E, Schneider ME. Trauma-induced systemic inflammatory response versus exercise-induced immunomodulatory effects. Sports Med 36: 373-384, 2006.
- 7. Vargas NT, Marino F. A neuroinflammatory model for acute fatigue during exercise. Sports Med 44: 1479-1487, 2014.
- Carmichael MD, Davis JM, Murphy EA, Carson JA, Van Rooijen N. Role of brain macrophages on IL-1beta and fatigue following eccentric exercise-induced muscle damage. Brain Behav Immun 24: 564-568, 2010.
- Straub RH. Insulin resistance, selfish brain, and selfish immune system: an evolutionarily positively selected program used in chronic inflammatory diseases. Arthritis Res Ther 16 Suppl 2: S4, 2014.
- 10. Mason S. Lactate Shuttles in Neuroenergetics-Homeostasis, Allostasis and Beyond. Front Neurosci 11: 43, 2017.
- Matsui T, Omuro H, Liu YF, Soya M, Shima T. Astrocytic glycogen-derived lactate fuels the brain during exhaustive exercise to maintain endurance capacity. Proc Natl Acad Sci U S A 114: 6358-6363, 2017.
- Robertson CV, Marino FE. Prefrontal and motor cortex EEG responses and their relationship to ventilatory thresholds during exhaustive incremental exercise. Eur J Appl Physiol 115: 1939-1948, 2015.
- Dworak M, Diel P, Voss S, Hollmann W, Strüder HK. Intense exercise increases adenosine concentrations in rat brain: implications for a homeostatic sleep drive. Neuroscience 150: 789-795, 2007.
- Zheng X, Hasegawa H. Administration of caffeine inhibited adenosine receptor agonist-induced decreases in motor performance, thermoregulation, and brain neurotransmitter release in exercising rats. Pharmacol Biochem Behav 140: 82-89, 2016.
- Hillion J, Canals M, Torvinen M, Casado V, Scott R. Coaggregation, cointernalization, and codesensitization of adenosine A2A receptors and dopamine D2 receptors. J Biol Chem 277: 18091-18097, 2002.
- Amann M. Significance of Group III and IV muscle afferents for the endurance exercising human. Clin Exp Pharmacol Physiol 39: 831-835, 2012.
- Hill AV, Lupton H. Muscular Exercise, Lactic Acid, and the Supply and Utilization of Oxygen. QJM os-16: 135-171, 1923.
- Brooks GA. Cell-cell and intracellular lactate shuttles. J Physiol 587: 5591-5600, 2009.
- 19. Garcia-Alvarez M, Marik P, Bellomo R. Sepsis-associated hyperlactatemia. Crit Care 18: 503, 2014.

- Kemppainen J, Aalto S, Fujimoto T, Kalliokoski KK, Långsjö J. High intensity exercise decreases global brain glucose uptake in humans. J Physiol 568: 323-332, 2005.
- 21. Schurr A. Lactate: the ultimate cerebral oxidative energy substrate? J Cereb Blood Flow Metab 26: 142-152, 2006.
- 22. Dienel GA. Brain lactate metabolism: the discoveries and the controversies. J Cereb Blood Flow Metab 32: 1107-1138, 2012.
- Morales-Alamo D, Losa-Reyna J, Torres-Peralta R, Martin-Rincon M, Perez-Valera M. What limits performance during whole-body incremental exercise to exhaustion in humans? J Physiol 593: 4631-4648, 2015.
- 24. Newsholme E, Acworth, IN, Bellotti P, Benzi, G, Ljungqvist, A. Amino acids, brain neurotransmitters and a functional link between muscle and brain that is important in sustained exercise. In World Symposium on Doping in Sport : Florence, Centro Affari, 10th-12th May, 1987; book exhibition catalogue, S. 25-. Rom: International Athletic Foundation, 1988.
- Cordeiro LM, Guimarães JB, Wanner SP, La Guardia RB, Miranda RM. Inhibition of tryptophan hydroxylase abolishes fatigue induced by central tryptophan in exercising rats. Scand J Med Sci Sports 24: 80-88, 2014.
- Strasser B, Geiger D, Schauer M, Gatterer H, Burtscher M, Fuchs D. Effects of Exhaustive Aerobic Exercise on Tryptophan-Kynurenine Metabolism in Trained Athletes. PLoS ONE 11: e0153617, 2016.
- Meeusen R, Watson P, Hasegawa H, Roelands B, Piacentini MF. Central fatigue: the serotonin hypothesis and beyond. Sports Med 36: 881-909, 2006.
- Taylor JL, Amann M, Duchateau J, Meeusen R, Rice CL. Neural Contributions to Muscle Fatigue: From the Brain to the Muscle and Back Again. Med Sci Sports Exerc 48: 2294-2306, 2016.
- Balthazar CH, Leite LH, Ribeiro RM, Soares DD, Coimbra CC. Effects of blockade of central dopamine D1 and D2 receptors on thermoregulation, metabolic rate and running performance. Pharmacol Rep 62: 54-61, 2010
- Morris G, Berk M, Walder K, Maes M. Central pathways causing fatigue in neuro-inflammatory and autoimmune illnesses. BMC Med 13: 28, 2015.
- Nakatomi Y, Mizuno K, Ishii A, Wada Y, Tanaka M. Neuroinflammation in Patients with Chronic Fatigue Syndrome/Myalgic Encephalomyelitis: An ¹¹C-(R)-PK11195 PET Study. J Nucl Med 55: 945-950, 2014.
- Patrick Neary J, Roberts AD, Leavins N, Harrison MF, Croll JC, Sexsmith JR. Prefrontal cortex oxygenation during incremental exercise in chronic fatigue syndrome. Clin Physiol Funct Imaging 28: 364-372, 2008.
- Smith LL. Cytokine hypothesis of overtraining: a physiological adaptation to excessive stress? Med Sci Sports Exerc 32: 317-331, 2000.
- Blankert JP. Neuroinflammation in burnout patients. In Neuroinflammation in burnout patients, Conference Proceedings, 2014.
- 35. Vargas NT, Marino F. A neuroinflammatory model for acute fatigue during exercise. Sports Med 44: 1479-1487, 2014.
- Gleeson M. Interleukins and exercise. The Journal of physiology, 529 Pt 1(Pt 1), 1, 2000.
- Robson-Ansley PJ, de Milander L, Collins, M, Noakes TD. Acute interleukin-6 administration impairs athletic performance in healthy, trained male runners. Can. J. Appl. Physiol. 29(4): 411-418, 2004.

- Pedersen BK, Febbraio MA. Muscles, exercise and obesity: skeletal muscle as a secretory organ. Nat Rev Endocrinol, 2012.
- Steensberg A, Febbraio MA, Osada T, Schjerling P, van Hall G. Interleukin-6 production in contracting human skeletal muscle is influenced by pre-exercise muscle glycogen content. J Physiol 537: 633-639, 2001.
- 40. Febbraio MA, Hiscock N, Sacchetti M, Fischer CP, Pedersen BK. Interleukin-6 is a novel factor mediating glucose homeostasis during skeletal muscle contraction. Diabetes 53: 1643-1648, 2004.
- 41. Steensberg A, Fischer CP, Keller C, Møller K, Pedersen BK. IL-6 enhances plasma IL-1ra, IL-10, and cortisol in humans. Am J Physiol Endocrinol Metab 285: E433-E437, 2003.
- 42. Hill EE, Zack E, Battaglini C, Viru M, Viru A, Hackney AC. Exercise and circulating Cortisol levels: The intensity threshold effect. J Endocrinol Invest 31: 587-591, 2008.
- 43. Carmichael MD, Davis JM, Murphy EA, Brown AS, Carson JA. Recovery of running performance following muscle-damaging exercise: relationship to brain IL-1beta. Brain Behav Immun 19: 445-452, 2005.
- 44. Carmichael MD, Davis JM, Murphy EA, Brown AS, Carson JA. Role of brain IL-1beta on fatigue after exercise-induced muscle damage. Am J Physiol Regul Integr Comp Physiol 291: R1344-R1348, 2006.
- 45. Zendedel A, Habib P, Dang J, Lammerding L, Hoffmann S. Omega-3 polyunsaturated fatty acids ameliorate neuroinflammation and mitigate ischemic stroke damage through interactions with astrocytes and microglia. J Neuroimmunol 278: 200-211, 2015.
- Shi Z, Ren H, Huang Z, Peng Y, He B. Fish Oil Prevents Lipopolysaccharide-Induced Depressive-Like Behavior by Inhibiting Neuroinflammation. Mol Neurobiol 54: 7327-7334, 2017.
- 47. Djordjevic B, Baralic I, Kotur-Stevuljevic J, Stefanovic A, Ivanisevic J. Effect of astaxanthin supplementation on muscle damage and oxidative stress markers in elite young soccer players. J Sports Med Phys Fitness 52: 382-392, 2012.
- Bruunsgaard H, Galbo H, Halkjaer-Kristensen J, Johansen TL, MacLean DA, Pedersen BK. Exercise-induced increase in serum interleukin-6 in humans is related to muscle damage. J Physiol 499 (Pt 3): 833-841, 1997.
- Simpson RJ, Cosgrove C, Chee MM, McFarlin BK, Bartlett DB. Senescent phenotypes and telomere lengths of peripheral blood T-cells mobilized by acute exercise in humans. Exerc Immunol Rev 16: 40-55, 2010.
- 50. Krüger K, Lechtermann A, Fobker M, Völker K, Mooren FC. Exercise-induced redistribution of T lymphocytes is regulated by adrenergic mechanisms. Brain Behav Immun 22: 324-338, 2008.
- 51. Fisher-Wellman K, Bloomer RJ. Acute exercise and oxidative stress: a 30 year history. Dyn Med 8: 1, 2009.
- 52. Fogarty MC, Hughes CM, Burke G, Brown JC, Trinick TR. Exercise-induced lipid peroxidation: Implications for deoxyribonucleic acid damage and systemic free radical generation. Environ Mol Mutagen 52: 35-42, 2011
- 53. Ostrowski K, Rhode T, Asp S, Schjerling P, Pedersen B. Pro□and anti□inflammatory cytokine balance in strenuous exercise in humans, 1999.
- 54. Gleeson M. Immune function in sport and exercise. J Appl Physiol 103: 693-699, 2007.

- 55. Heck TG, Schöler CM, de Bittencourt PI. HSP70 expression: does it a novel fatigue signalling factor from immune system to the brain? Cell Biochem Funct 29: 215-226, 2011.
- Bårdsen K, Nilsen MM, Kvaløy JT, Norheim KB, Jonsson G, Omdal R. Heat shock proteins and chronic fatigue in primary Sjögren's syndrome. Innate Immun 22: 162-167, 2016.
- 57. Marshall JC. The gut as a potential trigger of exercise-induced inflammatory responses. Can J Physiol Pharmacol 76: 479-484, 1998.
- Pals KL, Chang RT, Ryan AJ, Gisolfi CV. Effect of running intensity on intestinal permeability. J Appl Physiol 82: 571-576, 1997.
- 59. Snipe RMJ, Khoo A, Kitic CM, Gibson PR, Costa RJS. The impact of exertional-heat stress on gastrointestinal integrity, gastrointestinal symptoms, systemic endotoxin and cytokine profile. Eur J Appl Physiol 118: 389-400, 2018.
- Yeh YJ, Law LY, Lim CL. Gastrointestinal response and endotoxemia during intense exercise in hot and cool environments. Eur J Appl Physiol 113: 1575-1583, 2013.
- 61. Mutlu E, Keshavarzian A, Engen P, Forsyth CB, Sikaroodi M, Gillevet P. Intestinal dysbiosis: a possible mechanism of alcohol-induced endotoxemia and alcoholic steatohepatitis in rats. Alcohol Clin Exp Res 33: 1836-1846, 2009.
- 62. Shing CM, Peake JM, Lim CL, Briskey D, Walsh NP. Effects of probiotics supplementation on gastrointestinal permeability, inflammation and exercise performance in the heat. Eur J Appl Physiol 114: 93-103, 2014.
- 63. Brock-Utne JG, Gaffin SL, Wells MT, Gathiram P, Sohar E. Endotoxaemia in exhausted runners after a long-distance race. S Afr Med J 73: 533-536, 1988.
- 64. Lambert GP. Stress-induced gastrointestinal barrier dysfunction and its inflammatory effects. J Anim Sci 87: E101-E108, 2009.
- 65. Guo S, Al-Sadi R, Said HM, Ma TY. Lipopolysaccharide causes an increase in intestinal tight junction permeability in vitro and in vivo by inducing enterocyte membrane expression and localization of TLR-4 and CD14. Am J Pathol 182: 375-387, 2013.
- Dokladny K, Zuhl MN, Moseley PL. Intestinal epithelial barrier function and tight junction proteins with heat and exercise. J Appl Physiol 120: 692-701, 2016.
- 67. Shimada A, Hasegawa-Ishii S. Histological Architecture Underlying Brain-Immune Cell-Cell Interactions and the Cerebral Response to Systemic Inflammation. Front Immunol 8: 17, 2017.
- 68. Erickson MA, Dohi K, Banks WA. Neuroinflammation: a common pathway in CNS diseases as mediated at the bloodbrain barrier. Neuroimmunomodulation 19: 121-130, 2012.
- 69. Dantzer R, Heijnen CJ, Kavelaars A, Laye S, Capuron L. The neuroimmune basis of fatigue. Trends Neurosci 37: 39-46, 2014.
- 70. Bailey DM, Evans KA, McEneny J, Young IS, Hullin DA. Exercise-induced oxidative-nitrosative stress is associated with impaired dynamic cerebral autoregulation and bloodbrain barrier leakage. Exp Physiol 96: 1196-1207, 2011.
- Vitkovic L, Konsman JP, Bockaert J, Dantzer R, Homburger V, Jacque C. Cytokine signals propagate through the brain. Mol Psychiatry 5: 604, 2000.
- 72. Kapural M, Krizanac-Bengez Lj, Barnett G, Perl J, Masaryk T. Serum S-100beta as a possible marker of blood-brain barrier disruption. Brain Res 940: 102-104, 2002.

- 73. Koh SX, Lee JK. S100B as a marker for brain damage and blood-brain barrier disruption following exercise. Sports Med 44: 369-385, 2014.
- 74. Stålnacke BM, Tegner Y, Sojka P. Playing ice hockey and basketball increases serum levels of S-100B in elite players: a pilot study. Clin J Sport Med 13: 292-302, 2003.
- Papa L, Ramia MM, Edwards D, Johnson BD, Slobounov SM. Systematic review of clinical studies examining biomarkers of brain injury in athletes after sports-related concussion. J Neurotrauma 32: 661-673, 2015.
- Loane DJ, Kumar A. Microglia in the TBI brain: The good, the bad, and the dysregulated. Exp Neurol 275 Pt 3: 316-327, 2016.
- 77. Mollayeva T, Kendzerska T, Mollayeva S, Shapiro CM, Colantonio A, Cassidy JD. A systematic review of fatigue in patients with traumatic brain injury: the course, predictors and consequences. Neurosci Biobehav Rev 47: 684-716, 2014.
- Ponsford JL, Ziino C, Parcell DL, Shekleton JA, Roper M. Fatigue and sleep disturbance following traumatic brain injury – their nature, causes, and potential treatments. J Head Trauma Rehabil 27: 224-233, 2012.
- 79. Erickson MA, Banks WA. Cytokine and chemokine responses in serum and brain after single and repeated injections of lipopolysaccharide: multiplex quantification with path analysis. Brain Behav Immun 25: 1637-1648, 2011.
- Banks WA, Robinson SM. Minimal penetration of lipopolysaccharide across the murine blood-brain barrier. Brain Behav Immun 24: 102-109, 2010.
- Banks WA, Gray AM, Erickson MA, Salameh TS, Damodarasamy M. Lipopolysaccharide-induced blood-brain barrier disruption: roles of cyclooxygenase, oxidative stress, neuroinflammation, and elements of the neurovascular unit. J Neuroinflammation 12: 223, 2015.
- 82. Quan N, Stern EL, Whiteside MB, Herkenham M. Induction of pro-inflammatory cytokine mRNAs in the brain after peripheral injection of subseptic doses of lipopolysaccharide in the rat. J Neuroimmunol 93: 72-80, 1999.
- Black EAE, Cancelliere NM, Ferguson AV. Regulation of Nervous System Function by Circumventricular Organs, Neuroimmune Pharmacology. 25-37, 2017.
- 84. Vallières L, Rivest S. Regulation of the genes encoding interleukin-6, its receptor, and gp130 in the rat brain in response to the immune activator lipopolysaccharide and the proinflammatory cytokine interleukin-1beta. J Neurochem 69: 1668-1683, 1997.
- 85. Roth J, Harré EM, Rummel C, Gerstberger R, Hübschle T. Signaling the brain in systemic inflammation: role of sensory circumventricular organs. Front Biosci 9: 290-300, 2004.
- Chakravarty S, Herkenham M. Toll-like receptor 4 on nonhematopoietic cells sustains CNS inflammation during endotoxemia, independent of systemic cytokines. J Neurosci 25: 1788-1796, 2005.
- Nakano Y, Furube E, Morita S, Wanaka A, Nakashima T, Miyata S. Astrocytic TLR4 expression and LPS-induced nuclear translocation of STAT3 in the sensory circumventricular organs of adult mouse brain. J Neuroimmunol 278: 144-158, 2015.
- Furube E, Morita M, Miyata S. Characterization of neural stem cells and their progeny in the sensory circumventricular organs of adult mouse. Cell Tissue Res 362: 347-365, 2015.

- Thayer JF, Sternberg EM. Neural aspects of immunomodulation: focus on the vagus nerve. Brain Behav Immun 24: 1223-1228, 2010.
- Berthoud H-R, Neuhuber WL. Functional and chemical anatomy of the afferent vagal system. Auton Neurosci 85: 1-17, 2000.
- 91. Dantzer R, Heijnen CJ, Kavelaars A, Laye S, Capuron L. The neuroimmune basis of fatigue. Trends Neurosci 37: 39-46, 2014.
- 92. de La Serre CB, de Lartigue G, Raybould HE. Chronic exposure to low dose bacterial lipopolysaccharide inhibits leptin signaling in vagal afferent neurons. Physiol Behav 139: 188-194, 2015.
- 93. Fleshner M, Goehler LE, Schwartz BA, McGorry M, Martin D. Thermogenic and corticosterone responses to intravenous cytokines (IL-1beta and TNF-alpha) are attenuated by subdiaphragmatic vagotomy. J Neuroimmunol 86: 134-141, 1998.
- 94. Lavoie JM. The contribution of afferent signals from the liver to metabolic regulation during exercise. Can J Physiol Pharmacol 80: 1035-1044, 2002.
- 95. Pereira BC, Lucas G, da Rocha AL, Pauli JR, Ropelle ER. Eccentric Exercise Leads to Glial Activation but not Apoptosis in Mice Spinal Cords. Int J Sports Med 36: 378-385, 2015.
- Alvarez P, Levine JD, Green PG. Eccentric exercise induces chronic alterations in musculoskeletal nociception in the rat. Eur J Neurosci 32: 819-825, 2010.
- 97. Chiang CY, Sessle BJ, Dostrovsky JO. Role of astrocytes in pain. Neurochem Res 37: 2419-2431, 2012.
- Chacur M, Lambertz D, Hoheisel U, Mense S. Role of spinal microglia in myositis-induced central sensitisation: an immunohistochemical and behavioural study in rats. Eur J Pain 13: 915-923, 2009.
- 99. Louati K, Berenbaum F. Fatigue in chronic inflammation a link to pain pathways. Arthritis Res Ther 17: 254, 2015.
- 100. Maes M, Leunis JC. Normalization of leaky gut in chronic fatigue syndrome (CFS) is accompanied by a clinical improvement: effects of age, duration of illness and the translocation of LPS from gram-negative bacteria. Neuro Endocrinol Lett 29: 902-910, 2008.
- 101. Pal GD, Shaikh M, Forsyth CB, Ouyang B, Keshavarzian A, Shannon KM. Abnormal lipopolysaccharide binding protein as marker of gastrointestinal inflammation in Parkinson disease. Front Neurosci 9: 306, 2015.
- 102. Köhler CA, Maes M, Slyepchenko A, Berk M, Solmi M. The Gut-Brain Axis, Including the Microbiome, Leaky Gut and Bacterial Translocation: Mechanisms and Pathophysiological Role in Alzheimer's Disease. Curr Pharm Des 22: 6152-6166, 2016.
- 103. Berk M, Williams LJ, Jacka FN, O'Neil A, Pasco JA. So depression is an inflammatory disease, but where does the inflammation come from? BMC Med 11: 200, 2013.
- Gárate I, Garcia-Bueno B, Madrigal JL, Caso JR, Alou L. Stress-induced neuroinflammation: role of the Toll-like receptor-4 pathway. Biol Psychiatry 73: 32-43, 2013.
- 105. Arakawa H, Blandino P, Deak T. Central infusion of interleukin-1 receptor antagonist blocks the reduction in social behavior produced by prior stressor exposure. Physiol Behav 98: 139-146, 2009.
- 106. Vollmer-Conna U, Fazou C, Cameron B, Li H, Brennan C. Production of pro-inflammatory cytokines correlates with the symptoms of acute sickness behaviour in humans. Psychol Med 34: 1289-1297, 2004.

- 107. Puri BK, Holmes J, Hamilton G. Eicosapentaenoic acid-rich essential fatty acid supplementation in chronic fatigue syndrome associated with symptom remission and structural brain changes. Int J Clin Pract 58: 297-299, 2004.
- Lochmiller RL, Deerenberg C. Trade-offs in evolutionary immunology: just what is the cost of immunity? OIKOS 88: 87-99, 2000.
- McEwen BS. Allostasis and allostatic load: implications for neuropsychopharmacology. Neuropsychopharmacology 22: 108-124, 2000.
- Delmastro-Greenwood MM, Piganelli JD. Changing the energy of an immune response. Am J Clin Exp Immunol 2: 30-54, 2013.
- Orihuela R, McPherson CA, Harry GJ. Microglial M1/M2 polarization and metabolic states. Br J Pharmacol 173: 649-665, 2016.
- 112. Mason S, van Furth AM, Mienie LJ, Engelke UF, Wevers RA. A hypothetical astrocyte-microglia lactate shuttle derived from a 1H NMR metabolomics analysis of cerebrospinal fluid from a cohort of South African children with tuberculous meningitis. Metabolomics 11: 822-837, 2015.
- 113. Matsui T, Soya S, Kawanaka K, Soya H. Brain Glycogen Decreases During Intense Exercise Without Hypoglycemia: The Possible Involvement of Serotonin. Neurochem Res 40: 1333-1340, 2015.
- 114. Simpson RJ, Florida-James GD, Cosgrove C, Whyte GP, Macrae S. High-intensity exercise elicits the mobilization of senescent T lymphocytes into the peripheral blood compartment in human subjects. J Appl Physiol 103: 396-401, 2007.
- 115. Gleeson M. Immune function in sport and exercise. Edinburgh; New York: Churchill Livingstone Elsevier: British Association of Sport and Exercise Sciences, 2006.
- 116. Dimeloe S, Burgener AV, Grählert J, Hess C. T-cell metabolism governing activation, proliferation and differentiation; a modular view. Immunology 150: 35-44, 2017.
- 117. Brown FF, Bigley AB, Sherry C, Neal CM, Witard OC. Training status and sex influence on senescent T-lymphocyte redistribution in response to acute maximal exercise. Brain Behav Immun 39: 152-159, 2014.
- 118. Spielmann G, McFarlin BK, O'Connor DP, Smith PJ, Pircher H, Simpson RJ. Aerobic fitness is associated with lower proportions of senescent blood T-cells in man. Brain Behav Immun 25: 1521-1529, 2011.
- Bours MJ, Swennen EL, Di Virgilio F, Cronstein BN, Dagnelie PC. Adenosine 5'-triphosphate and adenosine as endogenous signaling molecules in immunity and inflammation. Pharmacol Ther 112: 358-404, 2006.
- Bajgar A, Kucerova K, Jonatova L, Tomcala A, Schneedorferova I. Extracellular adenosine mediates a systemic metabolic switch during immune response. PLoS Biol 13: e1002135, 2015.
- Dolezal, T. Adenosine: A selfish-immunity signal? Oncotarget
 6: 32307-32308, 2015
- 122. Boison D, Chen JF, Fredholm BB. Adenosine signaling and function in glial cells. Cell Death Differ 17: 1071-1082, 2010.
- 123. Cunha RA. Adenosine as a neuromodulator and as a homeostatic regulator in the nervous system: different roles, different sources and different receptors. Neurochem Int 38: 107-125, 2001.
- 124. Choi J, Tanaka K, Cao Y, Qi Y, Qiu J. Identification of a Plant Receptor for Extracellular ATP. Science 343: 290-294, 2014.

- 125. Gaikwad S, Patel D, Agrawal-Rajput R. CD40 Negatively Regulates ATP-TLR4-Activated Inflammasome in Microglia. Cell Mol Neurobiol 37: 351-359, 2017.
- 126. Beamer E, Gölöncsér F, Horváth G, Bekő K, Otrokocsi L. Purinergic mechanisms in neuroinflammation: An update from molecules to behavior. Neuropharmacology 104: 94-104, 2016.
- 127. Sperlágh B, Baranyi M, Haskó G, Vizi ES. Potent effect of interleukin-1 beta to evoke ATP and adenosine release from rat hippocampal slices. J Neuroimmunol 151: 33-39, 2004.
- 128. Saura J, Angulo E, Ejarque A, Casadó V, Tusell JM. Adenosine A2A receptor stimulation potentiates nitric oxide release by activated microglia. J Neurochem 95: 919-929, 2005.
- 129. Villalta SA, Nguyen HX, Deng B, Gotoh T, Tidball JG. Shifts in macrophage phenotypes and macrophage competition for arginine metabolism affect the severity of muscle pathology in muscular dystrophy. Hum Mol Genet 18: 482-496, 2009.
- 130. Shieh CH, Heinrich A, Serchov T, van Calker D, Biber K. P2X7-dependent, but differentially regulated release of IL-6, CCL2, and TNF- α in cultured mouse microglia. Glia 62: 592-607, 2014.
- Pocock JM, Kettenmann H. Neurotransmitter receptors on microglia. Trends Neurosci 30: 527-535, 2007.
- 132. Ferre S, Euler GV, Johansson B, Fredholm BB, Fuxe K. Stimulation of high-affinity adenosine A2 receptors decreases the affinity of dopamine D2 receptors in rat striatal membranes. Proc Natl Acad Sci U S A 88: 7238-7241, 1991.
- 133. Beggiato S, Tomasini MC, Borelli AC, Borroto-Escuela DO, Fuxe K. Functional role of striatal A2A, D2, and mGlu5 receptor interactions in regulating striatopallidal GABA neuronal transmission. J Neurochem 138: 254-264, 2016.
- 134. Correa M, Pardo M, Bayarri P, López-Cruz L, San Miguel N. Choosing voluntary exercise over sucrose consumption depends upon dopamine transmission: effects of haloperidol in wild type and adenosine A A AKO mice. Psychopharmacology 233: 393-404, 2016.
- 135. Zheng X, Hasegawa H. Administration of caffeine inhibited adenosine receptor agonist-induced decreases in motor performance, thermoregulation, and brain neurotransmitter release in exercising rats. Pharmacol Biochem Behav 140: 82-89, 2016.
- 136. Zheng X, Takatsu S, Wang H, Hasegawa H. Acute intraperitoneal injection of caffeine improves endurance exercise performance in association with increasing brain dopamine release during exercise. Pharmacol Biochem Behav 122: 136-143, 2014.
- 137. Roelands B, Buyse L, Pauwels F, Delbeke F, Deventer K, Meeusen R. No effect of caffeine on exercise performance in high ambient temperature. Eur J Appl Physiol 111: 3089-3095, 2011.
- Krüger K, Mooren FC. Exercise-induced leukocyte apoptosis. Exerc Immunol Rev 20: 117-134, 2014.
- Wang JS, Huang YH. Effects of exercise intensity on lymphocyte apoptosis induced by oxidative stress in men. Eur J Appl Physiol 95: 290-297, 2005.
- Turner JE, Bosch JA, Drayson MT, Aldred S. Assessment of oxidative stress in lymphocytes with exercise. J Appl Physiol 111: 206-211, 2011.
- 141. Rock KL, Kono H. The inflammatory response to cell death. Annu Rev Pathol 3: 99-126, 2008.

- 142. Goehler LE, Gaykema RP, Nguyen KT, Lee JE, Tilders FJ. Interleukin-1beta in immune cells of the abdominal vagus nerve: a link between the immune and nervous systems? J Neurosci 19: 2799-2806, 1999.
- 143. Margeli A, Skenderi K, Tsironi M, Hantzi E, Matalas AL. Dramatic elevations of interleukin-6 and acute-phase reactants in athletes participating in the ultradistance foot race spartathlon: severe systemic inflammation and lipid and lipoprotein changes in protracted exercise. J Clin Endocrinol Metab 90: 3914-3918, 2005.
- Biber K, Neumann H, Inoue K, Boddeke HW. Neuronal 'On' and 'Off' signals control microglia. Trends Neurosci 30: 596-602, 2007.
- 145. Haskó G, Pacher P, Vizi ES, Illes P. Adenosine receptor signaling in the brain immune system. Trends Pharmacol Sci 26: 511-516, 2005.
- Carman AJ, Mills JH, Krenz A, Kim DG, Bynoe MS. Adenosine receptor signaling modulates permeability of the bloodbrain barrier. J Neurosci 31: 13272-13280, 2011.
- 147. Calabrese EJ. Hormesis: principles and applications. Homeopathy 104: 69-82, 2015.
- 148. Radak Z, Ishihara K, Tekus E, Varga C, Posa A. Exercise, oxidants, and antioxidants change the shape of the bell-shaped hormesis curve. Redox Biol 12: 285-290, 2017.
- 149. Gimeno-Bayón J, López-López A, Rodríguez MJ, Mahy N. Glucose pathways adaptation supports acquisition of activated microglia phenotype. J Neurosci Res 92: 723-731, 2014.
- 150. Moreira TJ, Pierre K, Maekawa F, Repond C, Cebere A. Enhanced cerebral expression of MCT1 and MCT2 in a rat ischemia model occurs in activated microglial cells. J Cereb Blood Flow Metab 29: 1273-1283, 2009.
- 151. Robertson CV, Marino FE. Cerebral responses to exercise and the influence of heat stress in human fatigue. J Therm Biol 63: 10-15, 2017.
- 152. Juffermans NP, Verbon A, van Deventer SJ, Buurman WA, van Deutekom H. Serum concentrations of lipopolysaccharide activity-modulating proteins during tuberculosis. J Infect Dis 178: 1839-1842, 1998.
- 153. Peterson PK, Hu S, Sheng WS, Kravitz FH, Molitor TW. Thalidomide Inhibits Tumor Necrosis Factor-α Production by Lipopolysaccharide- and Lipoarabinomannan-Stimulated Human Microglial Cells. J Infect Dis 172: 1137-1140, 1995.
- 154. Wheeler DS, Chase MA, Senft AP, Poynter SE, Wong HR, Page K. Extracellular Hsp72, an endogenous DAMP, is released by virally infected airway epithelial cells and activates neutrophils via Toll-like receptor (TLR)-4. Respir Res 10: 31, 2009.

- 155. Vabulas RM, Ahmad-Nejad P, Ghose S, Kirschning CJ, Issels RD, Wagner H. HSP70 as endogenous stimulus of the Toll/interleukin-1 receptor signal pathway. J Biol Chem 277: 15107-15112, 2002.
- 156. Dalsgaard MK, Quistorff B, Danielsen ER, Selmer C, Vogelsang T, Secher NH. A reduced cerebral metabolic ratio in exercise reflects metabolism and not accumulation of lactate within the human brain. J Physiol 554: 571-578, 2004.
- 157. Mason S, Reinecke CJ, Kulik W, van Cruchten A, Solomons R, van Furth AM. Cerebrospinal fluid in tuberculous meningitis exhibits only the L-enantiomer of lactic acid. BMC Infect Dis 16: 251, 2016.
- 158. Hanff TC, Furst SJ, Minor TR. Biochemical and anatomical substrates of depression and sickness behavior. Isr J Psychiatry Relat Sci 47: 64-71, 2010.
- 159. Del Cerro, S., Borrell J. Interleukin-1 Affects the Behavioral Despair Response in Rats by An Indirect Mechanism Which Requires Endogenous CRF. Brain Research, 528: 162-164, 1990.
- 160. Holst SC, Landolt H-P. Sleep Homeostasis, Metabolism, and Adenosine. Current Sleep Medicine Reports 1: 27-37, 2015.
- Arrigoni E, Chamberlin NL, Saper CB, McCarley RW. Adenosine inhibits basal forebrain cholinergic and noncholinergic neurons in vitro. Neuroscience 140: 403-413, 2006.
- 162. Basheer R, Porkka-Heiskanen T, Strecker RE, Thakkar MM, McCarley RW. Adenosine as a biological signal mediating sleepiness following prolonged wakefulness. Biol Signals Recept 9: 319-327, 2000.
- 163. Shao W, Zhang SZ, Tang M, Zhang XH, Zhou Z. Suppression of neuroinflammation by astrocytic dopamine D2 receptors via α B-crystallin. Nature 494: 90-94, 2013.
- 164. Bjerring PN, Dale N, Larsen FS. Acute hyperammonemia and systemic inflammation is associated with increased extracellular brain adenosine in rats: a biosensor study. Neurochem Res 40: 258-264, 2015.
- Wilkinson DJ, Smeeton NJ, Watt PW. Ammonia metabolism, the brain and fatigue; revisiting the link. Prog Neurobiol 91: 200-219, 2010.
- Robertson CV, Marino FE. A role for the prefrontal cortex in exercise tolerance and termination. J Appl Physiol 120: 464-466, 2016.
- 167. St Clair Gibson A, De Koning JJ, Thompson KG, Roberts WO, Micklewright D. Crawling to the finish line: why do endurance runners collapse? Implications for understanding of mechanisms underlying pacing and fatigue. Sports Med 43: 413-424, 2013.